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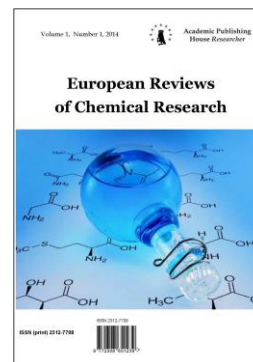
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Articles and Statements

Origin of Life in Hot Mineral Water from Hydrothermal Springs and Ponds. Effects of Hydrogen and Nascent Hydrogen. Analyses with Spectral Methods, pH and ORP

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Abstract

The studies were performed of the composition of water, its temperature, pH and oxidation reduction potential (ORP) value in experiments with modelling of primary hydrosphere and possible conditions for origin of first organic forms of life in hot mineral water in hydrothermal springs and open ponds. Experiments with hot mineral and seawater from Bulgaria by IR-spectroscopy with DNES-method and Thermo Nicolet Avatar 360 Fourier-transform IR were conducted. Cactus juice of *Echinopsis pachanoi* and Jellyfish *Aurelia aurita* from Black Sea were used as model systems. The reactions of condensation and dehydration in alkaline aqueous solutions with pH = 9–11, resulting in synthesis of larger organic molecules as polymers and short polipeptides from separate molecules, were considered and scrutinized. It was shown that hot alkaline mineral water with temperature from +65 °C to +95 °C and pH value from 8.5 to 10 and ORP with negative value is more suitable for the origination of life and living matter than other analyzed water samples. The pH value of seawater on the contrary is limited to the range of 7,5 to 8,4 units. The research was connected with estimation of the common local extremums in hot mineral and sea water, cactus juice and jellyfish.

Keywords: origin of life, hot mineral water, hydrothermal conditions, IR-spectroscopy.

1. Introduction

Previous biological experiments with D₂O and structural-conformational studies with deuterated molecules, enable to modeling conditions under which the first living forms of life might be evolved (Ignatov, Mosin, 2013). The content of deuterium in hot mineral water may be increased due to the physical chemical processes of the deuterium accumulation. It can be presumed that primary water might contain more deuterium at early stages of evolution of first living structures, and deuterium was distributed non-uniformly in the hydrosphere and atmosphere (Ignatov, Mosin, 2012). The primary reductive atmosphere of the Earth consisted basically of gas mixture CO, H₂, N₂, NH₃, CH₄, lacked O₂–O₃ layer protecting the Earth surface from rigid short-wave solar radiation carrying huge energy capable to cause radiolysis and photolysis of water.

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The point regards the influence of temperature on the processes in living matter. Recent studies have shown that the most favorable for the origin of life and living matter seem to be hot alkaline mineral waters interacting with CaCO_3 (Ignatov, 2010; Ignatov, Mosin, 2013). According to the law for conservation of energy the process of self-organization of primary organic forms in water solutions may be supported by thermal energy of magma, volcanic activity and solar radiation. According to J. Szostak, the accumulation of organic compounds in open lakes is more possible compared to the ocean (Szostak, 2011). Probably the life has begun near a hydrothermal vent: an underwater spout of hot water. Geothermal activity gives more opportunities for the origination of life. In 2009 A. Mulkidjanian and M. Galperin demonstrate that the cell cytoplasm contains potassium (K), zinc (Zn), manganese (Mn) and phosphate ions (P), which are not particularly widespread in the sea aquatorium (Mulkidjanian, Galperin, 2009). Colín-García and co-authors also summarize a set of experiments proposed to test the role of hydrothermal vents in prebiotic synthesis (Colín-García et al., 2016). B. Damer and D. Deamer have come to the conclusion that cell membranes cannot be formed in salty seawater. Before the continents formed, the only dry land on Earth would be volcanic islands, where rainwater would form ponds where lipids could form the first stages towards cell membranes. Only when true cells had evolved they gradually would adapt to saltier environments and enter the ocean (Damer, Deamer, 2015). J. Trevors and G. Pollack were proposed that the first cells on the Earth assembled in a hydrogel environment (Trevors, Pollack, 2005). Gel environments are capable of retaining water, oily hydrocarbons, solutes, and gas bubbles, and are capable of carrying out many functions, even in the absence of a membrane. Hydrocarbons are an organic compounds consisting entirely of hydrogen (H) and carbon (C). The analyses show the possible scenario of the syntheses of periodically molecules of life (Colón Santos, 2019). The data presented in this paper show that the origination of living matter most probably occurred in hot mineral water. This occurred in hydrothermal springs and ponds with hot mineral water. There had been possible also in hydrothermal vents in seawater with hot mineral water. An indisputable proof of this is the presence of stromatolites fossils. They lived in warm and hot water in zones of volcanic activity, which could be heated by magma and seem to be more stable than other first sea organisms (Ignatov, 2012).

The purpose of the research was studying the conditions of primary hydrosphere (temperature, pH, ORP, isotopic composition) for possible processes for origin of life and living matter in hot mineral water. There was studied primary atmosphere and interaction with hydrosphere and with effects of gas discharge. Various samples of water from Bulgaria were studied within the frames of the research.

2. Material and methods

2.1. Objects of Studying

The research by the IR-spectrometry with DNES-method (Antonov, 1990; Antonov, Ignatov, 1998) was carried out with samples of water taken from various water sources:

- 1 – mineral water (Rupite, Bulgaria);
- 2 – seawater (Varna, Bulgaria);

Sediments from hot mineral spring and pond in Rupite, Bulgaria and sea salt from Black Sea were studied using the Thermo Nicolet Avatar 360 Fourier-transform IR;

Cactus juice of *Echinopsis pachanoi* and Jellyfish *Aurelia aurita* (Varna, Bulgaria, Black Sea) were used as two model systems which were both investigated by the IR-spectrometry with DNES method.

2.2. IR-Spectroscopy

IR-spectra of water samples were registered on Thermo Nicolet Avatar 360 Fourier-transform IR (K. Chakarova) and Differential Non-equilibrium Spectrum (DNES).

2.3. pH indicator and oxidation reduction potential (ORP)

The research of pH and ORP with Hanna instruments was performed.

3. Results and discussion

Research of various samples of mineral water from mineral springs and seawater from Bulgaria was carried out. The hot mineral spring and ponds of Rupite are located in eastern foot of

the extinct volcano Kozhuh (Figure 1).



Fig. 1. Hot mineral ponds of Rupite, Bulgaria, foot of the extinct volcano Kozhuh

For this DNES method was employed for research of cactus juice *Echinopsis pachanoi* (Table 1). The cactus was selected as a model system because this plant contains approximately 90 % of water. The closest to the spectrum of cactus juice was the spectrum of mineral water contacting with Ca^{2+} and HCO_3^- ions from Rupite, Bulgaria (Table 1).

DNES-spectra of cactus juice and mineral water from Rupite, Bulgaria with HCO_3^- (1320–1488 mg/L), Ca^{2+} (29–36 mg/L) $t = 76\text{ }^\circ\text{C}$ (source) and $t = 52\text{--}54\text{ }^\circ\text{C}$ (open ponds depending the season) have magnitudes of local extremums at -0.1112 (11.15); -0.1187 (10.45); -0.1262 (9.83); -0.1287 (9.64) and -0.1387 eV (9.85 μm). Similar local extremum in the DNES-spectrum between cactus juice and seawater were detected at -0.1362 eV (9.10 μm).

Table 1. Results with DNES spectral method and Thermo Nicolet Avatar 360 Fourier-transform IR of cactus juice, jelly fish, sea water and salt, sea water and salt, mineral water and sediments from Rupite, Bulgaria

| -E (eV); λ (μm); k (cm^{-1}) DNES-method | -E (eV); λ (μm); k (cm^{-1}) DNES-method | λ (μm); k (cm^{-1}) Thermo Nicolet Avatar 360 Fourier-transform IR | -E (eV); λ (μm); k (cm^{-1}) DNES- method | λ (μm); k (cm^{-1}) Thermo Nicolet Avatar 360 Fourier- transform IR | -E (eV); λ (μm); k (cm^{-1}) DNES-method |
|--|--|--|--|--|--|
| Cactus juice | Mineral water Rupite (Bulgaria) | Mineral water Rupite (Bulgaria) | Sea Water Black Sea | Sea salt Black Sea | Jelly fish Black Sea |
| 0.1112 (11.15; 897) | 0.1112 (11.15; 897) | (11.44; 875) | | | |
| 0.1187 (10.45; 957) | 0.1187 (10.45; 957) | (10.95; 913) | | | 0.1200; (10.33; 968) |
| 0.1262 (9.82;1018) | 0.1262 (9.82; 1018) | | | | |
| 0.1287 (9.63; 1038) | 0.1287 (9.63; 1038) | (9.78; 1031) | | | |
| 0.1362 (9.10;1099) | – | (9.08;1101) | 0.1362 (9.10;1099) | | (0.1375; 9.02; 1109) |
| 0.1387 | 0.1387 (8.95; 1117) | – | | (8.93;1120) | |

Note: *The function of the distribution of energies Δf was measured in reciprocal electron volts (eV^{-1}). It is shown at which values of the spectrum $-E$ (eV) the biggest local maximums of this function are observed; λ – wave length; κ – wave number.

Common extremums in the IR-spectrum between cactus juice (DNES method) and minerals from the sediments (Thermo Nicolet Avatar 360 Fourier-transform IR) were detected at 11.44; 10.95; 9.78; 9.08; μm . Similar local extremum between cactus juice and sea salt was detected at 8.93 μm .

Study of the local extremums in DNES spectrum of jellyfish from Black Sea was performed (Table 1). There are two local extremums at 9.02 and 10.33 μm . The local extremum at 9.02 μm corresponds to the local extremums in sea salt (8.93 μm).

The local extremums of sediments from hot mineral spring in Rupite, Bulgaria were studied with Thermo Nicolet Avatar 360 Fourier-transform IR (Figure 2).

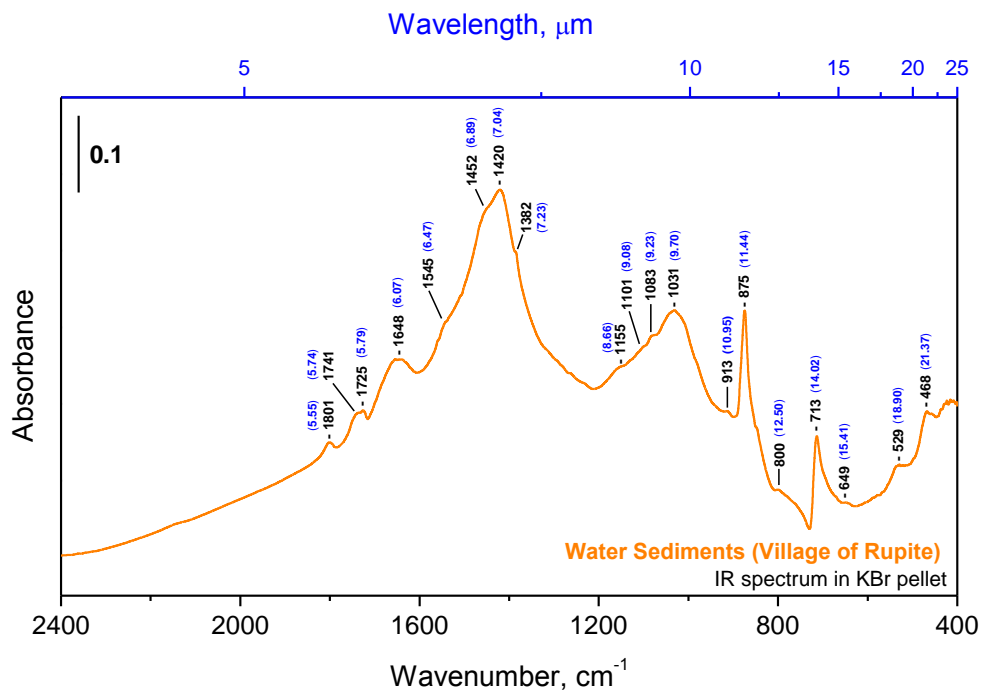


Fig. 2. IR-spectrum of water obtained from Rupite Village (Bulgaria)

The results with DNES method indicated that jellyfish *Aurelia aurita* had local extremums at 9.02 and 10.33 μm in IR-spectra (Table 1). Before measurements the jellyfish was kept in seawater for several days. For comparison seawater has a local extremum at 9.10 μm with DNES method and Sea salt 8.93 μm with Nicolet Avatar 360 Fourier-transform IR (Figure 3). Jellyfish contains approximately 97 (w/w) % of water and is the most unstable living organism compared to those ones that form stromatolites. The explanation for this is the smaller concentration of salts and, therefore, the smaller number of local extremums in the IR-spectrum in relation to seawater.

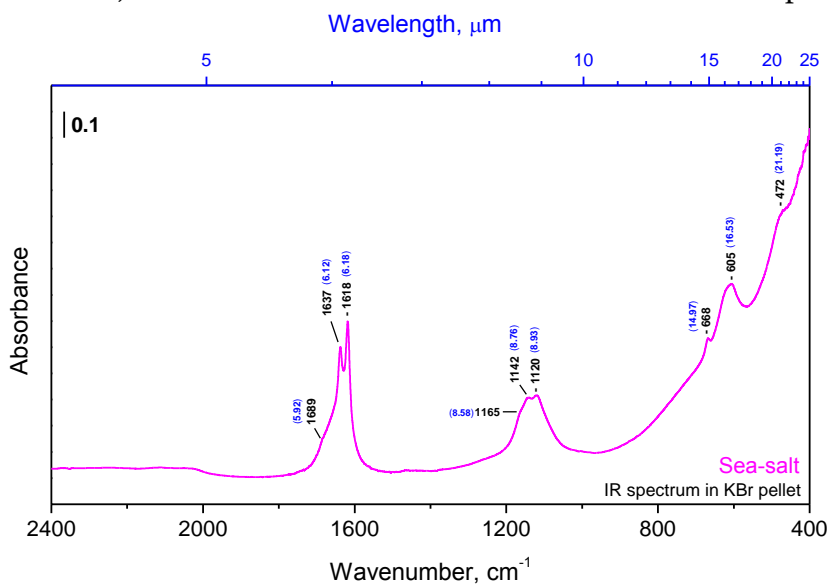
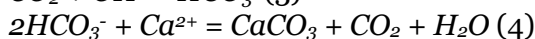
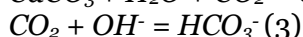
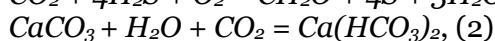


Fig. 3. IR-spectrum of seasalt obtained from Varna (Bulgaria)

Such a character of IR-spectrums and distribution of local extremums may prove that hot mineral alkaline water is preferable for origin and maintenance of life compared to other types of water analyzed by DNES and Nicolet Avatar 360 Fourier-transform IR. Thus, in hot mineral waters the local extremums in the IR-spectrums are more manifested compared to the local extremums obtained in IR-spectrum of the same water at a lower temperature (Ignatov, 2013). The difference in the local extremums in the interval from +20 °C to +95 °C at each 5 °C step is significant at $p < 0.05$ according to Student's t-criterion. These data indicate that the origination of life and living matter depends on the structure and physical chemical properties of water, as well as its temperature. The IR-spectrum of cactus juice is the closest to the IR- and DNES-spectrum of water, which contains bicarbonates and calcium ions typical for the formation of stromatolites. For this reason cactus juice was applied as a model system. The local extremums in IR-spectra of alkaline mineral water interacting with CaCO_3 and then seawater are the. The most closed closest to the local extremums in IR-spectrum of cactus juice.

In connection with these data the following reactions participating with CaCO_3 in aqueous solutions are important:



The equation (1) shows how some chemosynthetic bacteria use energy from the oxidation of H_2S and CO_2 to S and formaldehyde (CH_2O). The equation (2) is related to one of the most common processes in nature: in the presence of H_2O and CO_2 , CaCO_3 transforms into $\text{Ca}(\text{HCO}_3)_2$. In the presence of hydroxyl OH^- ions, CO_2 transforms into HCO_3^- (equation (3)). Equation (4) is valid for the process of formation of the stromatolites – the dolomite layered accretionary structures formed in shallow seawater by colonies of cyanobacteria. In 2010 D. Ward described fossilized stromatolites in the Glacier National Park (USA) (Schirber, 2010). Stromatolites aged 3.5 billion years had lived in warm and hot water in zones of volcanic activity, which could be heated by magma. This suggests that the first living forms evidently evolved in hot geysers (Pons et al., 2011). It is known that water in geysers is rich in carbonates, while the temperature is ranged from +60 to +100 °C and more than +100 °C. In 2011 a team of Japanese scientists under the leadership of T. Sugawara showed that life originated in warm or, more likely, hot water (Kurihara et al., 2011). DNA and synthetic enzymes created proto cells from aqueous solution of organic molecules. For this the initial solution was heated to a temperature close to water's boiling point +95 °C. Then its temperature was lowered to +65 °C with formation of proto cells with primitive membrane. This laboratory experiment is an excellent confirmation of the possibility that life originated in hot water.

The above-mentioned data can predict a possible transition from synthesis of small organic molecules under high temperatures to more complex organic molecules as proteins. There are reactions of condensation-dehydration of amino acids into separate blocks of peptides that occur under alkaline conditions, with $\text{pH} = 9\text{--}11$.

A research is conducted of Oxidation Reduction Potential (ORP) of hot mineral water from Rupite, Bulgaria. With temperature increase ORP gets reduced with (-70 mV) from 50 to 25° C (Table 2). The measured pH is 7.70. The change of ORP shows that in hot mineral water are released electrons in alkaline medium.

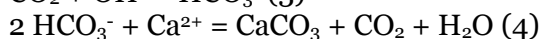
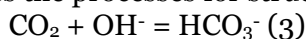
When reviewing the processes of life origin it is necessary to consider the composition of primary atmosphere 3.5 billion years ago. It contains H_2 , N_2 , CO_2 , CO. With the temperature rise of the water and boiling in the modern atmosphere, the bubbles contain oxygen and it gets acidified. The author suggests that the bubbles in the water in contact with the ancient atmosphere contain hydrogen, and the water gets more alkaline. Also ORP decreases and may result in negative values. An experiment is conducted with saturation of water from Rupite with hydrogen. The achieved average result is (-215 mV) with temperature 50°C.

Table 2. Results of temperature (°C) and Oxidation Reduction Potential (ORP) of mineral water from Rupite, Bulgaria

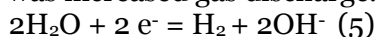
| Temperature (° C) | Oxidation Reduction Potential (ORP) (mV) |
|-------------------|--|
| 25 | 57 |
| 30 | 45 |
| 35 | 37 |
| 40 | 30 |
| 45 | 18 |
| 50 | -13 |

The difference is $(-215 \text{ mV}) - (-13 \text{ mV}) = (-202 \text{ mV})$. This indicates that in the ancient atmosphere were gained more electrons in the water. In such a way are achieved more hydroxyl groups (OH^-) and bicarbonate ions (HCO_3^-).

The interaction with calcium ions (Ca^{2+}) during exchange of electric charges as per formulae makes the processes for structuring of stromatolites by formulae (1) and (2) more active.

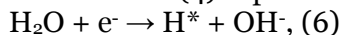


The following reaction (5) is valid in electrolysis. In the ancient atmosphere and hydrosphere there was increased gas discharge.

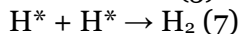


The same reaction contributed for the formation of stromatolites. Nowadays it is observed in electrolyzer devices for waters catholyte and anolyte. In the ancient hydrosphere and land the charge had been negative and in the atmosphere positive. The conditions had been optimal for "nature" electrolysis. In these direction had been the experiments of Miller ([Miller, 1953](#)).

The reaction (4) is possible to structure nascent hydrogen H^* ([Mehandjiev et al., 2019](#)).



The reaction (5) released in the gaseous form after recombination is formed:



The nascent hydrogen is very active for chemical reactions in primary hydrosphere for origin of life and additional formation of H_2 makes alkalizing effect.

The allocation of H_2O molecule when a peptide chain is formed is important factor in reaction of condensation of two amino acid molecules into dipeptide. As reaction of polycondensation of amino acids is accompanied by dehydration, the H_2O removal from reaction mixture speeds up the reaction rate. This testifies that formation of early organic forms may have occurred nearby active volcanoes, because at early periods of geological history volcanic activity occurred more actively than during subsequent geological times. However, dehydration accompanies not only amino acid polymerization, but also association of other small blocks into larger organic molecules, and also polymerization of nucleotides into nucleic acids. Such association is connected with the reaction of condensation, at which from one block a proton is removed, and from another – a hydroxyl group with the formation of H_2O molecule ([Ignatov, Mosin, 2012](#)).

The results with ORP show that free electrons in water support for amino acid polymerization and small blocks into larger organic molecules ([Ignatov, 2019](#)).

In 1969 the possibility of existence of condensation-dehydration reactions under conditions of primary hydrosphere was proven by M. Calvin ([Calvin, 1969](#)). From most chemical substances hydrocyanic acid (HCN) and its derivatives – cyanoamid (CH_2N_2) and dicyanoamid ($\text{HN}(\text{CN})_2$) possess dehydration ability and the ability to catalyze the process of linkage of H_2O from primary hydrosphere ([Mathews, Moser, 1968](#)). The presence of HCN in primary hydrosphere was proven by S. Miller's early experiments ([Miller, 1953](#)). Chemical reactions with HCN and its derivatives are complex with a chemical point of view; in the presence of HCN , CH_2N_2 and $\text{HN}(\text{CN})_2$ the condensation of separate blocks of amino acids accompanied by dehydration can proceed at normal temperatures in strongly diluted H_2O -solutions. These reactions show the results of synthesis from separate smaller molecules to larger organic molecules of polymers, e.g. proteins,

polycarboxydrates, lipids, and ribonucleic acids (Figure 4).

Furthermore, polycondensation reactions catalyzed by HCN and its derivatives depend on acidity of water solutions in which they proceed (Abelson, 1966). In acid aqueous solutions with pH = 4–6 these reactions do not occur, whereas alkaline conditions with pH = 9–11 promote their course. There has not been unequivocal opinion, whether primary water was alkaline, but it is probable that such pH value possessed mineral waters adjoining with basalts, i.e. these reactions could occur at the contact of water with basalt rocks, that testifies this hypothesis (Ignatov, Mosin, 2012).

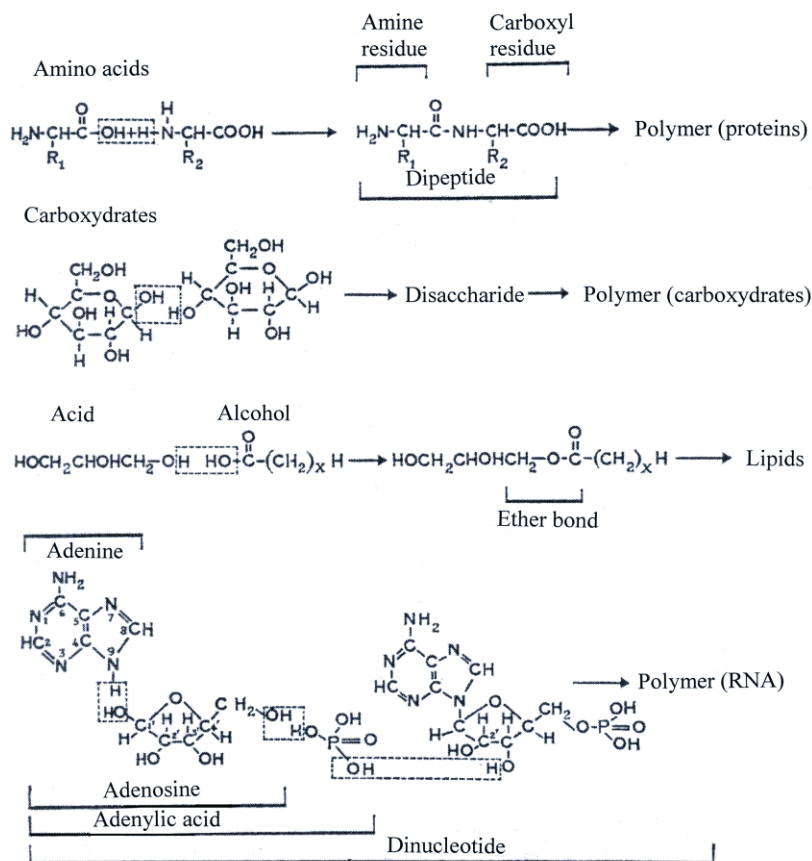
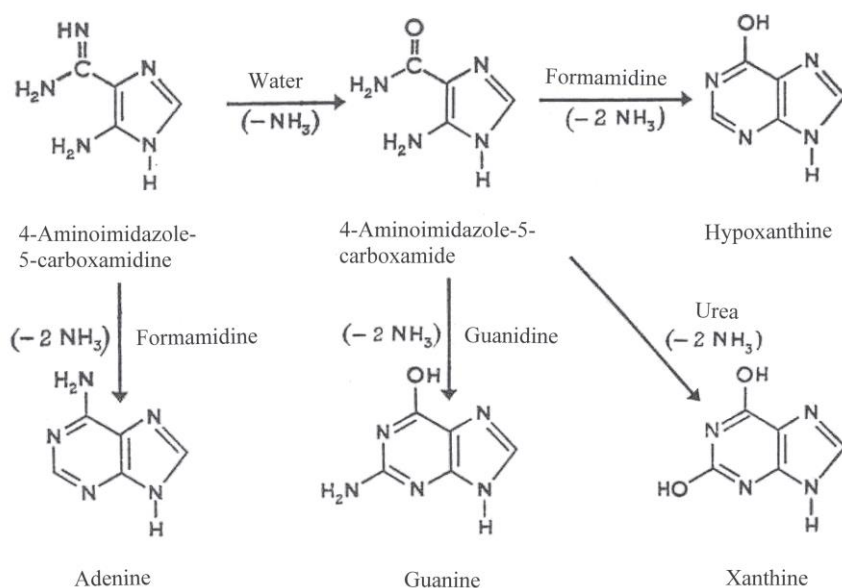


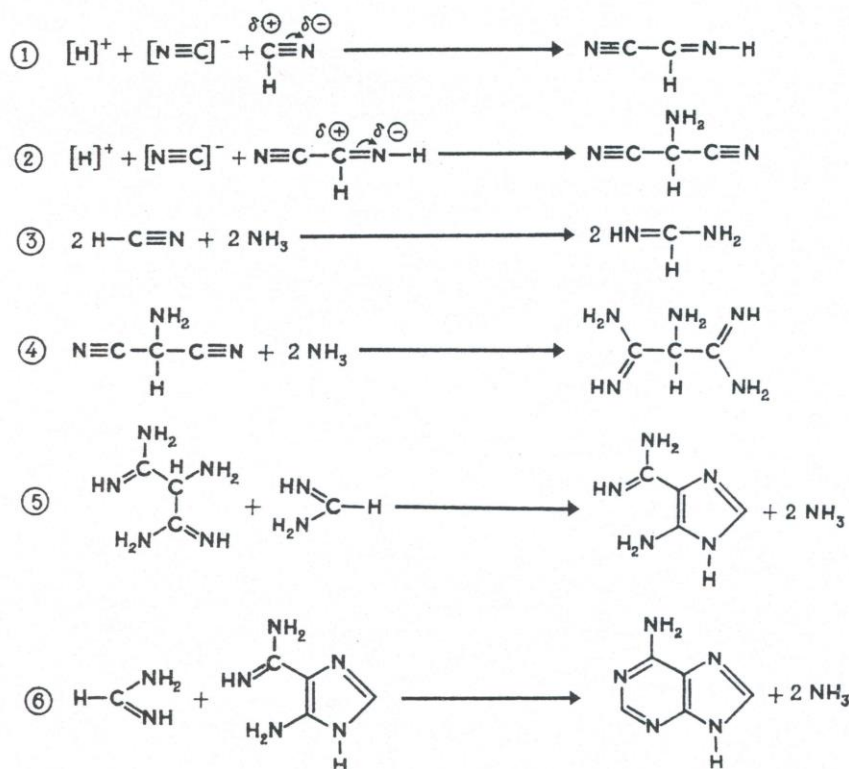
Fig. 4. Reactions of condensation and dehydration in alkaline conditions with pH = 9–11 catalyzed by HCN and its derivatives, resulting in synthesis from separate molecules larger organic molecules of polymers. The top three equations: condensation and the subsequent polymerization of amino acids in proteins; carbohydrates – in polycarboxydrates and acids and ethers – into lipids. The bottom equation – condensation of adenine with ribose and H_3PO_4 , leading to formation of dinucleotide

It should be noted, that geothermal sources might be used for synthesis of various organic molecules. Thus, amino acids were detected in solutions of formaldehyde CH_2O with hydroxylamine NH_2OH , formaldehyde with hydrazine (N_2H_4) in water solutions with HCN, after heating of a reactionary mixture to +95 °C (Harada, Fox, 1964). In model experiments reaction products were polymerized into peptide chains that are the important stage towards inorganic synthesis of protein. Purines and pyrimidines were formed in a reactionary mixture of water solution with a $\text{HCN}-\text{NH}_3$ (Figure 5). In other experiments amino acid mixtures were subjected to influence of temperatures from +60 °C up to +170 °C with formation of short protein-like molecules resembling early evolutionary forms of proteins subsequently designated as thermal proteinoids (Ignatov, Mosin, 2012). They consisted of 18 amino acids usually occurring in protein hydrolyzates. The synthesized proteinoids are similar to natural proteins on a number of other important properties, e.g. on linkage by nucleobases and ability to cause the reactions similar to those catalyzed by enzymes in living organisms as decarboxylation, amination, deamination, and oxidoreduction. Proteinoids are capable to catalytically decompose glucose (Fox, Krampitz, 1964) and

to have an effect similar to the action of α -melanocyte-stimulating hormone (Fox, Wang, 1968). The best results on polycondensation were achieved with the mixes of amino acids containing aspartic and glutamic acids, which are essential amino acids occurring in all modern living organisms.



a)



b)

Fig. 5. Prospective mechanisms of thermal (+95 °C) synthesis of purines in aqueous solutions: a) – synthesis of hypoxanthine, adenine, guanine and xanthine from 4-aminoimidazole-5-carboxamide, 4-aminoimidazole-5-carboxamide, water, NH_3 , formamidine and urea; b) – synthesis of adenine from NH_3 and HCN (total reaction: $5\text{HCN} = \text{adenine}$)

In natural conditions water was heated by the magma. The structure formed from heated water was evidently a result of self-organization. Living organisms are complex self-organizing systems. They are open because they constantly exchange substances and energy with the environment and change the entropy (Ignatov, 2011). The changes in the open systems are relatively stable in time. The stable correlation between components in an open system is called a dissipative structure. According to I. Prigozhin, the formation of dissipative structures and the elaboration to living cells is related to changes in entropy (Nikolis, Prigozhin, 1979).

The initial stage of evolution, apparently, was connected with formation of the mixtures of amino acids and nitrogenous substances – analogues of nucleic acids at high temperature. Such synthesis is possible in aqueous solutions under thermal conditions in the presence of H_3PO_4 . The next stage is polycondensation of amino acids into thermal proteinoids at temperatures 65–95 °C. After that membrane like structures were formed in a mix of proteinoids in hot water solutions. In 2011 T. Sugawara (Japan) created membrane like proto cells from aqueous solution of organic molecules, DNA and synthetic enzymes under temperature close to water's boiling point +95 °C (Sugawara, 2011).

4. Conclusion

The data obtained testify that origination of life and living matter depends on physical-chemical properties of water and external factors – temperatures, pH, ORP, electric discharges and isotopic composition. Hot mineral alkaline water interacting with $CaCO_3$ is closest to these conditions. Next in line with regard to quality is seawater. For chemical reaction of dehydration-condensation to occur in hot mineral water, water is required to be alkaline with pH range 9–11 and negative ORP. In warm and hot mineral waters the local extremums in IR-spectrums from 8 to 14 μm were more expressed in comparison with the local extremums measured in the same water samples with lower temperature. The new achievement is connected with chemical composition of ancient atmosphere and alkalization of the water from the hydrogen.

The research for origin of life and living matter in hot mineral water in mineral springs and open ponds (Ignatov, 2010; Szostak, 2011; Damer, Deamer, 2015) and in hydrothermal vents in the ocean (Mulkiđjanian, Galperin, 2009) give the possibilities for life at the surface of the moons of the planets in Solar System. According to the law for conservation of energy the process of self-organization of primary organic forms in water solutions may be supported by thermal energy of magma (Ignatov, 2010). The candidates are – Europa (moon of Jupiter), Titan and Enceladus (moons of Saturn).

5. Acknowledgements

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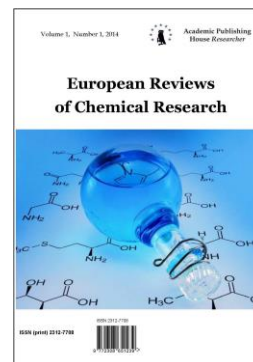
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Results with IR Spectroscopy of CortiNon+ on the Development of Experimental *Graffi* Tumor on Hamsters

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Abstract

Studies were conducted with product CortiNon+. CortiNon+ is mix of Liquid Progesterone/DHEA. Progesterone is progestin, a female hormone, produced mostly in the ovaries.

DHEA is an endogenous hormone secreted by the adrenal gland of both males and females. The purpose of the research was to analyze the effects over blood serum of hamsters with *Graffi* tumor (Toshkova, 1995). The analyses have been conducted with Nonequilibrium Energy Spectrum (NES) and Differential Nonequilibrium Energy Spectrum (DNES) methods (Antonov, 1992; Ignatov, 1998). Experiments were carried out about the influence on tumor cells of mice in water. Reduction of DNES spectrum according to the control sample of cells in healthy animals was observed. (Antonov, 1992). Reduction has also been observed in DNES spectrum in blood serum of people having oncological diseases, compared to the one of healthy people (Ignatov, 2012). Such a reduction is most prevalent in (-0.1387 eV; 8.95 μm; 1117 cm⁻¹). The investigation of blood serum from hamsters injected with *Graffi* tumor cells and treated with CortiNon+ demonstrated a reduction in DNES spectrum in the range (-0.08 – -0.14 eV) (8.9 – 15.5 μm) (645–1129 cm⁻¹) (Toshkova et al., 2019).

Keywords: CortiNon+, *Graffi* tumor, NES, DNES.

1. Introduction

The research demonstrates the effects of CortiNon+ on the *Graffi* tumor of hamsters. The hamsters are separated into 5 groups noted as Gr. 1, Gr. 2, Gr. 3, Gr. 4 and Gr. 5. The first two groups Gr.1 and Gr.2 are for studying the effects of CortiNon+ on the tumor development. Gr. 1 includes hamsters that started taking CortiNon+ once daily for 7 days before the injection of *Graffi* tumor cells and then continued taking daily it until the end of the experiment. Gr. 2 is from hamsters that were s.c. injected with the same amount of tumor cells and started taking CortiNon+ on the same day as the tumor cell injection. The other groups are used as control. Gr. 3 is of non-treated tumor-bearing hamsters. Gr. 4 is of CortiNon+ treated healthy hamsters, and Gr. 5 is of healthy non-treated animals.

The research is conducted using spectral methods NES and DNES (Antonov, 1990; Antonov, Ignatov, 1998).

The spectrum analyses with methods NES and DNES are conducted on the 10th day after transplantation of *Graffi* tumor cells, which coincides with the appearance (formation) of

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subcutaneous tumor in the trial animals.

2. Materials and Methods

Experimental design. All the animals were divided into 5 groups as follows.

Gr. 1 – The hamsters from this group started taking CortiNon+ once daily for 7 days before the injection with 5×10^4 *Graffi* tumor cells per hamster in the back area, and then continued taking it once daily until the end of the experiment.

Gr. 2 – These hamsters were s.c. injected with the same amount of tumor cells as Gr. 1 and started taking CortiNon+ on the same day of the tumor injection.

Gr. 3 – Hamsters with *Graffi* tumor as a control untreated group.

Gr. 4 – Healthy hamsters taking CortiNon+ as a control group.

Gr. 5 – Healthy hamsters as a control untreated group.

1. Experimental animals

Hamsters, breed “Golden Syrian”, aged 2-4 months with weight around 90-100 g were used in the trials. The animals were grown in standard conditions in individual plastic cages with free access to food and water.

2.2. Experimental tumor model

Tumor cells ($1-2 \cdot 10^6$) from the experimental *Graffi* solid tumor are transplanted subcutaneously in the back of hamsters. Between days 7 and 15 after the transplantation tumor appears, grows progressively and the hamsters die around 30-35 days. In this tumor model 100 % tumor transplantation and 100 % mortality are observed. No spontaneous tumor's regression takes place (Toshkova, 1995).

2.3. Ethical aspects

All experiments were conducted in accordance with the European convention for protection of vertebrate animals, used for experimental and other scientific purposes (OJ L 222) and approved from the National Veterinary Medical Office.

2.4. NES and DNES Spectral Analyses

The device for DNES spectral analysis based on an optical principle was designed by A. Antonov. For this, a hermetic camera for evaporation of water drops under stable temperature (+22–24 °C) conditions was used. The water drops were placed on a water-proof transparent pad, which consisted of thin maylar folio and a glass plate. The light was monochromatic with filter for yellow color with wavelength at $\lambda = 580 \pm 7$ nm. The device measures the angle of evaporation of water drops from 72.3° to 0° . The DNES-spectrum was measured in the range of -0.08 – -0.1387 eV or $\lambda = 8.9$ – 13.8 μm using a specially designed computer program. The main estimation criterion in these studies was the average energy ($\Delta E_{\text{H...O}}$) of hydrogen O...H-bonds between H_2O molecules in water samples and hamster serum blood.

3. Results and discussion

3.1. Parameters of NES and DNES spectrums of CortiNon+

The spectrum analysis is conducted on the 10th day after transplantation of *Graffi* tumor cells, which coincides with the appearance (formation) of subcutaneous tumor in the trial animals. The energy spectrum of water is characterized by a non-equilibrium process of water droplets evaporation; therefore, the term non-equilibrium spectrum (NES) of water is used.

The difference $\Delta f(E) = f(\text{samples of water}) - f(\text{control sample of water})$ – is called the “differential non-equilibrium energy spectrum of water” (DNES). The Figure 1 shows that on the X-axis are depicted three scales. The energies of hydrogen bonds among H_2O molecules are calculated in electronvolts (eV). On the Y-axis is depicted the function of distribution of H_2O molecules according to energies $f(E)$, measured in reciprocal electronvolts unit eV^{-1} . The local extremums of water samples are detected at $E = -0.1112$ eV, $E = -0.1212$ eV and $E = -0.1387$ eV. The value measured at $E = -0.1212$ eV is characteristic for anti-inflammatory effect (Ignatov et al., 2014) The value measured at $E = -0.1112$ eV is characteristic for the presence of Ca^{2+} ions in water

(Antonov, 1995). The value measured at $E = -0.1387$ eV is characteristic for inhibiting the growth of tumor cells (Ignatov, Mosin, 2012). Experiments conducted by Antonov with cancer cells of mice in water demonstrated a reduction of this local extremum to a negative value in DNES spectra.

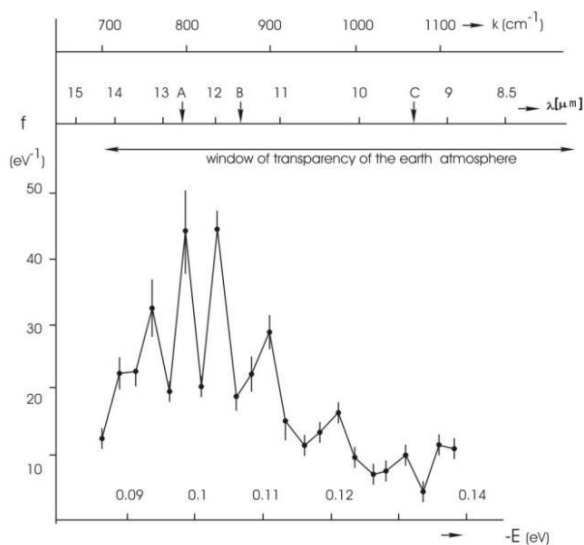


Fig. 1. The NES-spectrum of deionized water (chemical purity – 99.99 %; pH – 6,0–7,5; electric conductivity – 10 $\mu\text{S}/\text{cm}$): the horizontal axis shows the energy of the H...O hydrogen bonds in the associates ($-E$ (eV)); the vertical axis – the energy distribution function – f (eV^{-1}); k – the vibration frequency of the H–O–H atoms (cm^{-1}); λ – wavelength (μm)

The following results of the effects of CortiNon+ with NES and DNES methods are obtained:

3.1.1. The difference of DNES spectra between Gr. 1 and Gr. 3 is

$\Delta E = (-0.1194 \text{ eV}) - (-0.1156 \text{ eV}) = -3.8 \pm 1,1 \text{ meV}$. The difference is essential and shows effect of CortiNon+ on *Graffi* tumor of molecular and cell level

3.1.2. The difference of DNES spectra between Gr. 1 and Gr. 5 is

$\Delta E = (-0.1194 \text{ eV}) - (-0.1179 \text{ eV}) = -1.5 \pm 1,1 \text{ meV}$. This result is achieved for the products with anti tumor effect.

3.1.3. The difference of DNES spectra between Gr. 1 and Gr. 4 is

$\Delta E = (-0.1194 \text{ eV}) - (-0.1177 \text{ eV}) = -1.7 \pm 1,1 \text{ meV}$. This result is achieved for the products with anti tumor effect. For Gr. 1 CortiNon+ with implanted *Graffi* tumor after 7 days the result is higher according to healthy hamsters from Gr. 4 with CortiNon+.

3.1.4. The difference of DNES spectra between DNES spectrum of CortiNon+ and control sample of deionized water is $\Delta E = (-0.1230 \text{ eV}) - (-0.1083 \text{ eV}) = -14.7 \pm 1,1 \text{ meV}$. This is the maximal effect of the product for structuring of hydrogen bonds among water molecules.

3.2. The mathematical models of CortiNon+

The mathematical models of CortiNon+ of Gr.1, Gr.3 and Gr.5 give valuable information for the possible number of hydrogen bonds as percent of H_2O molecules with different values of distribution of energies (Table 1 and Fig. 2). These distributions are basically connected with the restructuring of H_2O molecules having the same energies.

The average energy ($E_{\text{H...O}}$) of hydrogen H...O- bonds among H_2O molecules of the samples of blood serum of hamsters was measured for the following groups.

Gr. 1 – The result of NES for Gr.1 is $E = -0.1194 \text{ eV}$.

Gr. 2 – The result of NES for Gr. 2 is $E = -0.1168 \text{ eV}$.

Gr. 3 – The result of NES for Gr. 3 is $E = -0.1156 \text{ eV}$.

Gr. 4 – The result of NES for Gr. 4 is $E = -0.1177 \text{ eV}$.

Gr. 5 – The result of NES for Gr. 5 is $E = -0.1179 \text{ eV}$.

The Table 1 and Figure 2 show the mathematical Models of CortiNon+ for Gr. 1, Gr. 3 and Gr. 5.

Table 1. Mathematical Models of CortiNon+ for Gr. 1, Gr. 3 and Gr. 5

| -E(eV) x-axis | 1 st group CortiNon+ 7 days before <i>Graffi</i> tumor (%((-E _{value})* (-E _{total value}) ^{**}) | 3 ^d group <i>Graffi</i> tumor (%((-E _{value})* (-E _{total value}) ^{**}) | 5 th group Healthy hamsters (%((-E _{value})* (-E _{total value}) ^{**}) | -E(eV) x-axis | 1 st group CortiNon+ 7 days before <i>Graffi</i> tumor (%((-E _{value})* (-E _{total value}) ^{**}) | 3 ^d group <i>Graffi</i> tumor (%((-E _{value})* (-E _{total value}) ^{**}) | 5 th group Healthy hamsters (%((-E _{value})* (-E _{total value}) ^{**}) |
|------------------|--|---|--|------------------|--|---|--|
| 0.0937 | 0 | 0 | 0 | 0.1187 | 4.5 | 9.7 | 9.5 |
| 0.0962 | 0 | 0 | 0 | 0.1212 | 13.6 | 6.1 | 0 |
| 0.0987 | 0 | 5.1 | 4.7 | 0.1237 | 0 | 9.7 | 0 |
| 0.1012 | 9.1 | 5.1 | 4.7 | 0.1262 | 0 | 5.1 | 9.5 |
| 0.1037 | 9.1 | 9.7 | 9.5 | 0.1287 | 9.1 | 0 | 0 |
| 0.1062 | 9.1 | 9.7 | 19.4 | 0.1312 | 4.5 | 5.1 | 0 |
| 0.1087 | 0 | 5.1 | 4.7 | 0.1337 | 4.5 | 5.1 | 9.5 |
| 0.1112 | 18.1 | 5.1 | 0 | 0.1362 | 4.5 | 0 | 9.5 |
| 0.1137 | 0 | 9.7 | 0 | 0.1387 | 13.9 | 0 | 9.5 |
| 0.1162 | 0 | 9.7 | 9.5 | - | - | | - |

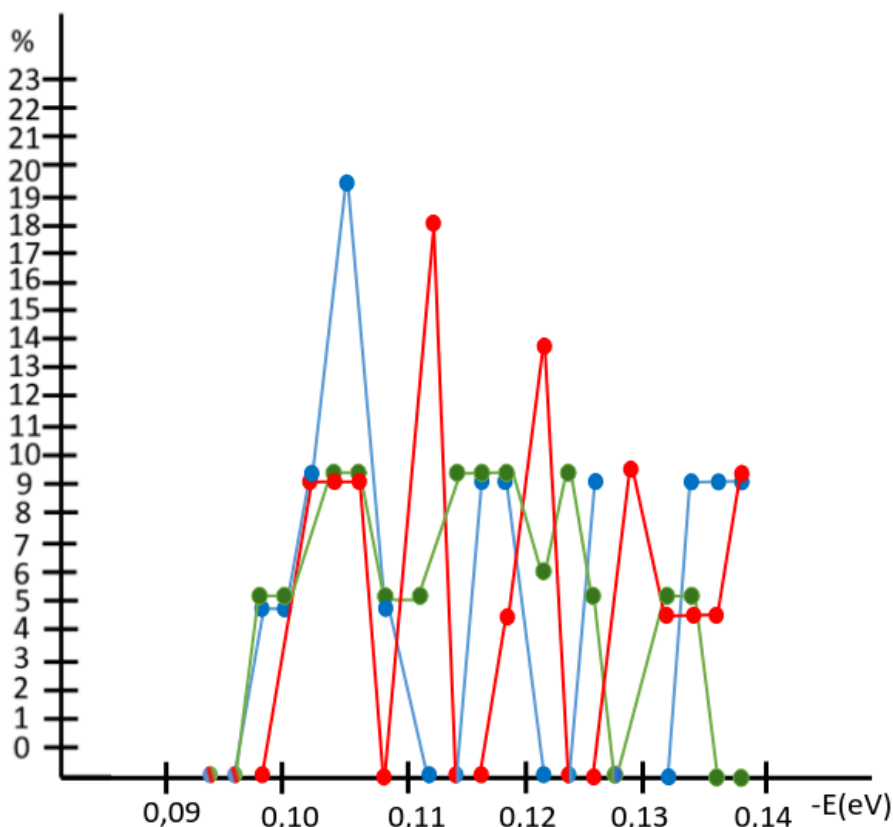


Fig. 2. Mathematical Models of CortiNon+ for Gr. 1, Gr. 3 and Gr. 5

Notes:

E = -0.1112 eV is the local extremum for stimulating effect on nervous system and improvement of nervous conductivity.

$E = -0.1212$ eV is the local extremum for anti inflammatory effect.

$E = -0.1387$ eV is the local extremum for inhibition of development of tumor cells of molecular level.

* ($-E_{\text{value}}$) stands for the value of hydrogen bonds energy for one parameter of ($-E$).

** ($-E_{\text{total value}}$) stands for the total value of hydrogen bonds energy.

Figure 2 shows the distribution (% , ($-E_{\text{value}}/(-E_{\text{total value}})$) of H_2O molecules of CortiNon+ of Gr. 1, Gr. 3 and Gr. 5 respectively.

$E = -0.1112$ eV is the local extremum for stimulating effect on nervous system and improvement of nervous conductivity. The effect of CortiNon+ is 18.1 % for Gr.1, 5.1 % for Gr. 3 and 0 % for Gr. 5.

$E = -0.1212$ eV is the local extremum for anti inflammatory effect. The effect is 13.6 % for Gr. 1, 6.1 % for Gr. 3 and 0 % for Gr. 5.

$E = -0.1387$ eV is the local extremum for inhibition of development of tumor cells of molecular level. The effect is 13.9 % for Gr. 1, 0 % for Gr. 3 and 9.5 % for Gr. 5.

4. Conclusion

The influence of CortiNon+ on the development of *Graffi* tumor implanted in hamsters is assessed. The results show restructuring of water molecules in clusters with distribution that influences beneficially hamster (and human) health on molecular and cellular level.

The results from the application of NES and DENS spectrums and mathematical models confirm the effects of CortiNon+ on the nervous and endocrine systems, and anti-inflammatory and anti-tumor effects, as well. One interesting observation is that higher effect from the application of CortiNon+ on the tumor-bearing hamsters from Gr. 1 that have been treated 7 days before the injection is obtained relative to even the result of healthy hamsters from Gr. 4.

Product Name: CortiNon+

Inventor: Georgi D. Dinkov.

Trademark: Submitted to USPTO (USA) on December 10, 2017.

Product Distribution: IdeaLabs, LLC, 1200 18th Street NW #700, Washington, DC, USA.

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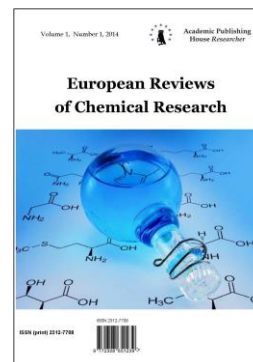
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Physicochemical and Microbiological Characteristics of Thermal Healing Spring Waters in the District of Burgas

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Abstract

Defined are the physicochemical properties of healing thermal spring waters in the area of Burgas District. It is shown that according to 18 controlled parameters included in the research, the thermal healing spring water village of Shivarovo, thermal healing spring water village of Polyanovo, fulfill the required conditions for drinking water.

The spring waters from the given four water sources are characterized by microbiological indicators, as the pathogenic micro-organisms are defined by the membrane method. It is established that thermal healing spring water Burgas Mineral baths, thermal healing spring water village of Shivarovo, thermal healing spring water of village of Polyanovo, fulfill the standard requirements. The healing water of village Judge, District of Burgas does not conform to the physicochemical indicators given for fluorides, and microbiological indicators with regards to coliform bacteria, *Escherichia coli* and enterococci.

Keywords: spring water, drinking water, physicochemical properties, microbiological indicators.

1. Introduction

In Bulgaria there are mineral and spring waters, which are not subjected to physicochemical and microbiological control by the Regional Health Inspectorate and microbiological control by they are the most use springs for drinking from the population. Similar springs are located in the territory of Haskovo District, Stara Zagora District, Varna District (Valcheva et al., 2013).

Although water is an unfavorable environment for the development of microorganisms and for the development of microorganisms, studies by many authors including heir, own, that microorganisms with valuable properties (enzymes, antibiotics, thermophile can acidophilic strains) are in mineral and non – thermal spring waters. This was proved by the results obtained from the experimental work carried out to determine the microflora of medicinal and spring waters in Haskovo, Stara Zagora, Plovdiv (Tumbariski et al., 2014) and Varna region (Valcheva, Ignatov, 2019).

Isolated bacteria from the healing and spring regions have been identified by *Bacillus subtilis*, *Bacillus cereus*, *Bacillus thuringiensis*, *Bacillus methylotrophicus*, *Aeromonashydrophila*.

The isolated bacteria from the healing and spring waters in the Plovdiv region have been identified by molecular genetic methods such as *Aeromonassobria*, *Klebsiellaoxytoca*, *Bacillus*

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amyloliquefaciens subsp. *plantarum*, *Bacillus thuringiensis*, *Bacillus cereus* (Valcheva et al., 2013, 2014).

Strains with high proteolytic, lipolytic and amylolytic activity were selected (Valcheva et al., 2013, 2014).

Antimicrobial activity of the strains of *Bacillus* sp., against the saprophytic and pathogenic microorganisms was detected: *Penicillium* sp., *Fusarium moliniforme*, *Rhizopus* sp., *Aspergillus niger*, *Aspergillus oryzae*, *Aspergillus awamori*, *Mucor* sp. *Enterococcus faecalis*, in the process of development and growth of the four *Bacillus* – *Bacillus cereus*, *Bacillus thuringiensis*, *Bacillus subtilis*, *Bacillus methylotrophicus* are the most active strains – *Bacillus methylotrophicus* PY5, *Bacillus cereus* LH1, *Bacillus cereus* WIF15 and *Bacillus thuringiensis* B62 (Valcheva et al., 2013, 2014).

Pathogenic bacteria exhibit resistance and 4 retain their vitality in the process of development and interaction between them and the strains of *Bacillus* sp. and at 37 °C.

A relatively low bactericidal effect was demonstrated against the (Gr+) bacterium *Enterococcus faecalis*. The isolated strains are likely to have a higher inhibitory ability against (Gr-) bacteria compared to (Gr+) bacteria (Valcheva et al., 2013, 2014).

The yeasts used in the genus *Candida* exhibit a simulating effect of two of *Bacillus* sp. – *Bacillus methylotrophicus* PY5, and *Bacillus cereus* LH1. This indicates that synergism has occurred between these microorganisms (Valcheva et al., 2013, 2014).

Mineral springs Burgas city

Burgas is the second largest seaside town in Bulgaria, located on the Southern Black Sea coast. In addition to the beautiful sea and spa – pious beaches, Burgas offers great opportunities for balneological treatment with mineral water, characteristic sea mud and lye.

This is one of the oldest balneological centers in Bulgaria. Mineral water is suitable for the treatment and prevention of diseases of the musculoskeletal system, peripheral nervous system, chronic gastritis and pyelonephritis, infertility, gout.

Water is also beneficial for strengthening the general state of the body.

Mineral spring District Sudievo

Sudievo is a village in southeastern Bulgaria. It is located in Aytos municipality. Water helps diseases of urinary system, disorders of locomotory system, endocrine diseases. It is suitable for daily use as drinking water. This water in Sudievo is hydrocarbonate, sodium, but contains fluoride. According to the requirements for drinking water, not mineral water, water should contain not more than 1.5 milligrams per liter of fluoride. The water in Sudievo contains much more in quantity than this chemical element. What those who consume this water need to know is that excessive ingestion of this fluoride per day can accordingly damage tooth enamel in young children. In the northern part of Aytosko Polje there are several mineral springs along the fault line: the "Smelly Fountain" near the village of Shivarovo and those near the villages of Cherry, Yabulchevo and Saedinie. Geothermal water with a flow rate of 30 L/s and a temperature of 51 °C emerges from deep drilling in the village of Polyanovo, which flows freely without being used. Analyses show that the sources have extremely good healing properties.

Medicinal properties of water: in diseases of the locomotory system, gastrointestinal, liver-bile and renal diseases.

2. Materials and methods

In the work are used thermal healing waters from the district of Burgas – thermal healing spring Burgas Mineral baths with water temperature of 41 °C, thermal healing spring village of Shivarovo with water temperature of 47 °C, thermal healing spring village of Polyanovo with water temperature of 51 °C, thermal healing spring village of Judge with water temperature of 51 °C.

2.1. Nutrient media

Nutrient agar (MPA) with contents (in %) – meat water, peptone – 1 %, agar – agar – 2 %. Endo's Medium (for defining of *Escherichia coli* and coliform bacteria) with contents (g/dm³) – peptone – 5,0; triptone – 5,0; lactose – 10,0; Na₂SO₃ – 1,4; K₂HPO₄ – 3,0; fuchsine – 0,14; agar – agar – 12,0, pH 7,5 – 7,7.

Nutrient gelatine (MPD) (for defining of *Pseudomonas aeruginosa*) with contents (in %) – Peptic digest of animal tissue; 25 % gelatin; pH = 7, 0-7, 2.

Medium for defining of enterococci (esculin – bile agar).

Medium for defining of sulphite reducing bacteria (Iron Sulphite Modified Agar).

Wilson-Bleer medium (for defining of sulphite reducing spore anaerobes (*Clostridium perfringens*) with contents(g/dm³) – 3 % Nutrient agar; 100 cm³ 20 % solution Na₂SO₃; 50 cm³ 20 % glucose solution; 10 cm³ 8 % solution of Fe₂SO₄.

2.2. Methods for analysis

Methods for physicochemical analysis

Method for determination of color according to Rublyovska Scale – method by Bulgarian State Standard (BDS) 8451: 1977;

Method for determination of smell at 20°C – method BDS 8451: 1977 technical device – glass mercury thermometer, conditions N^o 21;

Method for determination of turbidity – EN ISO 7027, technical device turbidimeter type TURB 355 IR ID N^o 200807088;

Method for determination of pH – BDS 3424: 1981, technical device pH meter type UB10 ID N^o UB10128148;

Method for determination of oxidisability – BDS 3413: 1981;

Method for determination of chlorides – BDS 3414: 1980;

Method for determination of nitrates – Validated Laboratory Method (VLM) – NO₃ – N^o 2, technical device photometer, “NOVA 60 A” ID N^o 08450505;

Method for determination of nitrites – VLM NO₃ – N^o 3, technical device photometer “NOVA 60 A” ID N^o 08450505;

Method for determination of ammonium ions – VLM – NH₄ – N^o 1, technical device photometer “NOVA 60 A” ID N^o 08450505;

Method for determination of general hardness – BDS ISO 6058;

Method for determination of sulphates – VLM – SO₄ – N^o 4, technical device photometer, “NOVA 60 A” ID N^o 08450505;

Method for determination of calcium – BDS ISO 6058;

Method for determination of magnesium – BDS 7211: 1982;

Method for determination of phosphates – VLM - PO₄ – N^o 5, technical device photometer, “NOVA 60 A” ID N^o 08450505;

Method for determination of manganese – VLM – Mn – N^o 7, technical device photometer, “NOVA 60 A” ID N^o 08450505;

Method for determination of iron – VLM – Fe – N^o 6, technical device photometer “NOVA 60 A” ID N^o 08450505;

Method for determination of fluorides – VLM – F – N^o 8, technical device photometer “NOVA 60 A” ID N^o 08450505;

Method for determination of electrical conductivity – BDS EN 27888, technical device – conductivity meter inoLabcond 720 ID N^o 11081137.

2.3. Methods for determination of microbiological indicators

Methods for evaluation of microbiological indicators according to Ordinance N^o 9/2001, Official State Gazette, issue 30, and decree N^o 178/23.07.2004 about the quality of water, intended for drinking purposes.

Method for determination of *Escherichia coli* and coliform bacteria – BDS EN ISO 9308 – 1: 2004;

Method for determination of enterococci – BDS EN ISO 7899 – 2;

Method for determination of sulphite reducing spore anaerobes – BDS EN 26461 – 2: 2004;

Method for determination of total number of aerobic and facultative anaerobic bacteria – BDS EN ISO 6222: 2002;

Method for determination of *Pseudomonas aeruginosa* – BDS EN ISO 16266 : 2008.

Determination of coli – titre by fermentation method – Ginchev’s method

Determination of coli – bacteria over Endo’s medium – membrane method.

Determination of sulphite reducing anaerobic bacteria (*Clostridium perfringens*) – membrane method.

3. Results and discussion

It is done a comparative physicochemical analysis of mineral spring waters at the territory of Burgas District by the main indicators (colour according to Rublyovska Scale, smell at 20°C, turbidity, pH, oxidisability, chlorides, nitrates, nitrites, ammonium ions, general hardness, sulphates, calcium, magnesium, phosphates, manganese, iron, fluorides, electrical conductivity). The results from these examinations are given in Table 1.

The trial data reveal that thermal healing spring water village of Shivarovo, thermal healing spring water village of Polyanovo swimming pool are in compliance with the controlled parameters set out in Ordinance N^o9/2001, Official State Gazette, issue 30, and decree N^o 178/23.07.2004 about the quality of water, intended for drinking purposes(RZI (Regional Health Inspection) – Burgas).

Table 1. Comparison of the examined spring waters in Burgas District by physicochemical properties

| Controlled parameter | Measuring unit | Maximum Limit Value | Result Burgas Mineral baths | Result Shivarovo | Result Polyanovo | Result Judge |
|--|-----------------------------------|-------------------------|-----------------------------|-------------------------|-------------------------|-------------------------|
| 1. Color according to Rublyovska Scale | Chromaticity Values | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers |
| 2. Smell at 20°C | Rating | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers |
| 3. Turbidity | NTU | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers | Acceptable to consumers |
| 4. pH indicator | pH values | ≥ 6,5 и ≤ 9,5 | 9,95 | 9 | 9,11 | 9,1 |
| 5. Oxidisability | mgO ₂ /dm ³ | 5,0 | 0,50 | 0,4 | 0,5 | 0,5 |
| 6. Chlorides | mg/ dm ³ | 250 | 30,7 | 26,3 | 26,3 | 26,0 |
| 7. Nitrates | mg/ dm ³ | 50 | 0,2 | 2,10 | 0,1 | 0,15 |
| 8. Nitrites | mg/ dm ³ | 0,50 | 0,007 | 0,00 | 0,006 | 0,005 |
| 9. Ammonium ions | mg/ dm ³ | 0,50 | 0,111 | 0,150 | 0,154 | 0,158 |
| 10. General hardness | mgekv/ dm ³ | 12 | 0,4 | 0,4 | 0,4 | 0,4 |
| 11. Sulphates | mg/ dm ³ | 250 | 37 | 34 | 35 | 36 |
| 12. Calcium | mg/ dm ³ | 150 | 120 | 118 | 116 | 117 |
| 13. Magnesium | mg/ dm ³ | 80 | 68 | 66 | 67 | 66 |
| 14. Phosphates | mg/ dm ³ | 0,5 | 0,015 | 0,016 | 0,016 | 1,018 |
| 15. Manganese | mg/ dm ³ | 50 | 0,0005 | 0,0008 | 0,0007 | 0,0009 |
| 16. Iron | µg/ dm ³ | 200 | 0,0016 | 0,0020 | 0,0022 | 0,0037 |
| 17. Fluorides | mg/ dm ³ | 1,5 | 7,73 | 0,4 | 1,48 | 5,5 |
| 18. Electrical conductivity | µS/ dm ³ | 2000 | 633 | 620 | 612 | 615 |

For the same spring waters are determined their microbiological indicators by the membrane method. In Table 2 are shown the experimental studies from the determination of total number of mesophilic aerobic and facultative anaerobic bacteria.

Table 2. Determination of total number of mesophilic aerobic and facultative anaerobic bacteria

| Examined water source | Indicator, cfu/cm ³ |
|---|--------------------------------|
| 1. Thermal healing spring Burgas Mineral baths with water temperature of 41°C | 6 ± 1 |
| 2. Thermal Healing Spring village of Shivarovowith water temperature of 41 °C | 11 – 17 |
| 3. Thermal Healing Springvillage of Polyanovowith water temperature of 51°C | 5 – 8 |
| 4. Thermal Healing Springvillage of Judge with water temperature of 51 °C | 120 – 150 |

According to the standard requirements from the examined water samples from the four springs, the water is clean.

The presence of coliforms and *Escherichia coli* is determined by the membrane method, and according to Ginchev's method. The trial results (Table 3 and Table 4) reveal that thermal healing spring Burgas Mineral baths with water temperature of 41°C, thermal healing spring village of Shivarovo with water temperature of 41°C, thermal healing spring village of Polyanovo with water temperature of 51°C swimming pool, are in compliance with the requirements for presence of coli bacteria. Thermal healing spring village of Polyanovo does not comply with the requirements for presence of coliform bacteria and enterococci. The given results are also confirmed by the analyses via the membrane method (Table 4). All the remaining indicators are determined by the membrane method.

Table 3. Coli – titre of thermal healing spring waters

| Name of water source | Coli – titre | Culture volumes 50cm ³ | Culture volumes 10cm ³ | Culture volumes 10cm ³ | Culture volumes 10cm ³ | Culture volumes 10cm ³ | Culture volumes 10cm ³ |
|--|--------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|
| 1. Thermal healing spring Burgas Mineral baths with water temperature of 41°C | > 100 | – | – | – | – | – | – |
| 2. Thermal Healing Spring village of Shivarovo with water temperature of 41 °C | > 100 | – | – | – | – | – | – |
| 3. Thermal Healing Springvillage of Polyanovo with water temperature of 51°C | > 100 | – | – | – | – | – | – |
| 4. Thermal Healing Spring village of Judge with water temperature of 51 °C | 80 | + Acid | + Acid | + Acid and gas | + Acid and gas | + Acid and gas | – |

Table 4. Microbiological indicators of spring waters in Burgas District

| Indicators | Measuring unit | Thermal healing spring Burgas Mineral baths with water temperature of 41°C | Thermal healing springvillage of Shivarovo with water temperature of 41 °C | Thermal healing spring village of Polyanovo with water temperature of 51 °C | Thermal healing springvillage of Judge with water temperature of 51 °C |
|------------------------|---------------------|--|--|---|--|
| Coliforms | cfu/cm ³ | 0/100 | 0/100 | 0/100 | 10/100 |
| <i>Escherichiacoli</i> | cfu/cm ³ | 0/100 | 0/100 | 0/100 | 10/100 |
| Enterococci | cfu/cm ³ | 0/100 | 8/100 | 0/100 | 8/100 |

| | | | | | |
|--|---------------------|-------|-------|-------|-------|
| Sulphite reducing anaerobic bacteria(<i>Clostridium perfringens</i>) | cfu/cm ³ | 0/100 | 0/100 | 0/100 | 0/100 |
| <i>Pseudomonas aeruginosa</i> | cfu/cm ³ | 0/250 | 0/250 | 0/250 | 0/250 |

Based on the conducted physicochemical and microbiological evaluations it is established that from the four examined springs at the territory of Burgas District by physicochemical parameters only thermal healing spring water village of Shivarovo, thermal healing spring water village of Polyanovo swimming pool correspond to all controlled parameters according to Ordinance № 9/2001, Official State Gazette, issue 30, and decree № 178 / 23.07.2004 about the quality of water, intended for drinking purposes. With regards to microbiological parameters thermal healing water Burgas Mineral baths, thermal healing spring village of Shivarovo, thermal healing spring water village of Polyanovo swimming pool are in compliance with the requirements for drinking water.

4. Conclusion

The research shows the effects of hot mineral water from Burgas region, Bulgaria.

There are the results with

- Comparison of the examined spring waters in Burgas District by physicochemical properties;
- Determination of total number of mesophilic aerobic and facultative anaerobic bacteria;
- Coli – titre of thermal healing spring waters;
- Microbiological indicators of spring waters in Burgas District.

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Standards

Ordinance №9/2001, Official State Gazette, issue 30.

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BDS8451: 1977 – defining of colour according to Rublyovska Scale, determination of smell at 20 °C.

EN ISO 7027 – determination of turbidity.

BDS3424: 1981 – determination of pH.

BDS3413: 1981 – determination of oxidisability.

BDS3414: 1980 – determination of chlorides.

BDS ISO 6058 – determination of calcium, determination of general hardness.

BDS EN 27888 – determination of electrical conductivity.

VLM – NH₄ – № 1 – determination of ammonium ions.

VLM – NO₃ – № 2 – determination of nitrates.

VLM – NO₂ – № 3 – determination of nitrites.

VLM – SO₄ – № 4 – determination of sulphates.

VLM – PO₄ – № 5 – determination of phosphates.

VLM – Fe – № 6 – determination of iron.

VLM – Mn – № 7 – determination of manganese.

VLM – F – № 8 – determination of fluorides.

BDS 7211: 1982 – determination of magnesium.

BDS EN ISO 7899 – 2 – determination of nitrates.

BDS EN ISO 9308 – 1: 2004 – determination of *Escherichia coli* and coliform bacteria.

BDS EN ISO 26461 – 2: 2004 – determination of sulphite reducing anaerobic bacteria (*Clostridium perfringens*).

BDS EN ISO 16266 – determination of *Pseudomonas aeruginosa*.

BDS EN ISO 7899 – 2 – determination of enterococci.

BDS EN ISO 6222: 2002 – determination of total number of aerobic and facultative anaerobic bacteria.